



# **IPHC 5-year Biological and Ecosystem Science Research Program: Update**

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***Biological and Ecosystem Science  
Branch Manager***

IPHC Interim Meeting

28 – 29 November, 2017



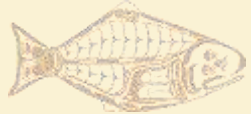
INTERNATIONAL PACIFIC  
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# Outline of the presentation



- **Update on the research activities of the Biological and Ecosystem Science Program**
- **Outcome of external funding applications**
- **Proposed research projects for 2018**
- **Establishment of the IPHC research laboratory**

Paper IPHC-2017-IM093-11



# Outline of the presentation



- **Update on the research activities of the Biological and Ecosystem Science Program**
- Outcome of external funding applications
- Proposed research projects for 2018
- Establishment of the IPHC research laboratory

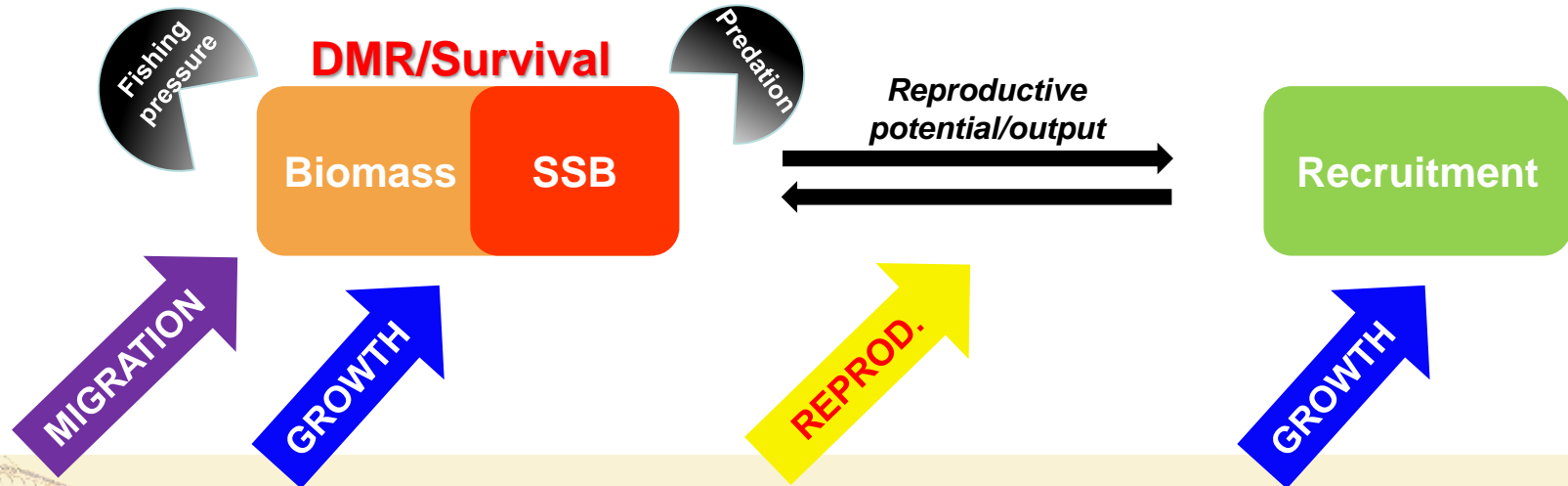


# Primary research activities at IPHC



## Primary objectives

- Identify and address *critical knowledge gaps* in the biology of the Pacific halibut
- Understand the influence of *environmental conditions* on halibut biology
- Apply resulting knowledge to reduce *uncertainty* in current stock assessment models



# Primary research areas at IPHC



1. **Reproduction**
  - SEX RATIO OF COMMERCIAL CATCH
  - IMPROVED MATURATION ESTIMATES OF SPAWNING BIOMASS
2. **Growth**
  - CHANGES IN SIZE AT AGE/BIOMASS
  - TOOLS TO ASSESS FISH CONDITION
3. **DMRs and post-release survival assessment**
  - BYCATCH SURVIVAL ESTIMATES
4. **Migration**
  - ADULT FEEDING AND REPRODUCTIVE MIGRATION
  - LARVAL DISPERSAL
5. **Genetics and genomics**
  - GENETIC STRUCTURE OF THE POPULATION
  - GENOMIC TOOLS (e.g. GENOME)



# Update of research activities at IPHC



## 1. Reproduction

- SEX RATIO OF COMMERCIAL CATCH
- IMPROVED MATURATION ESTIMATES OF SPAWNING BIOMASS

## 2. Growth

## 3. DMRs and post-release survival assessment

## 4. Migration

## 5. Genetics and genomics



# 1. Reproduction

**There are important knowledge gaps on the reproductive biology of the species**

- **SEX RATIO OF COMMERCIAL CATCH**
- **IMPROVED MATURATION ESTIMATES OF SPAWNING BIOMASS**

## **Projects:**

- 1. Sex marking and identification of genetic sex (Projects 621.15 and 621.16)***
- 2. Full characterization of the annual reproductive cycle (Project 674.11)***
- 3. Identification of genetic reproductive markers***

## **Objectives:**

- **Identification of genetic markers of sex and information on sex ratios.**
- **Knowledge on reproductive development, maturation, fecundity, environmental and hormonal control of reproduction.**
- **Scientific-based criteria to identify reproductive status and potential.**
- **Updated estimates of age and size at maturation.**
- **Information on skipped spawning.**



# 1. Reproduction

- **Development of genetic markers for sex identification (Project 621.16)**

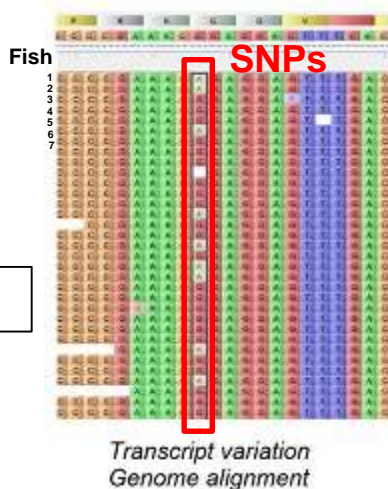
**Objectives:** To identify genetic markers for sex identification in the commercial catch

- Restriction site-associated DNA sequencing (RADseq) approach
  - 95 fish (55 female and 40 male) sequenced
  - 40,308 loci identified
  - 56 loci linked to sex (70 SNPs) based on  $F_{ST}$  values  $\geq 0.4$
  - Most females heterozygous and most males homozygous (ZW/ZZ)
  - 3 loci found only in females (0 only in males)
  - TaqMan assays developed for 2 sex-linked loci: Hs10183, Hs23885

Genetic assay accuracy (based on 199 morphologically sexed fish): 97.5%



Dr. Dan Drinan  
Dr. Lorenz Hauser



Drinan, Loher and Hauser (2017) *J. Heredity*. In Press





# 1. Reproduction

- ***Sex marking at sea and validation of genetic sex (Project 621.15)***

**Objectives:** To establish a method for physically marking sex by the commercial fleet and validation of marking efficiency by genetic sex identification



Dorsal Cut (Female)    Gill Plate Cut (Male)

2017

Reg Area	Sampled trips	Number of fish
2A	36	70
2B	5	84
2C	16	116
3A	10	113
3B	9	292
4A	2	77
4B	2	95
4C	4	86
4D	1	19
<b>TOTAL</b>	<b>84</b>	<b>929</b>

- 2016 (Reg Area 2B; 10 R/V; 288 samples)

79% marking accuracy (validated genetically)

- 2017 (Coastwide; 87 R/V; 929 samples)

??% marking accuracy (validated genetically)

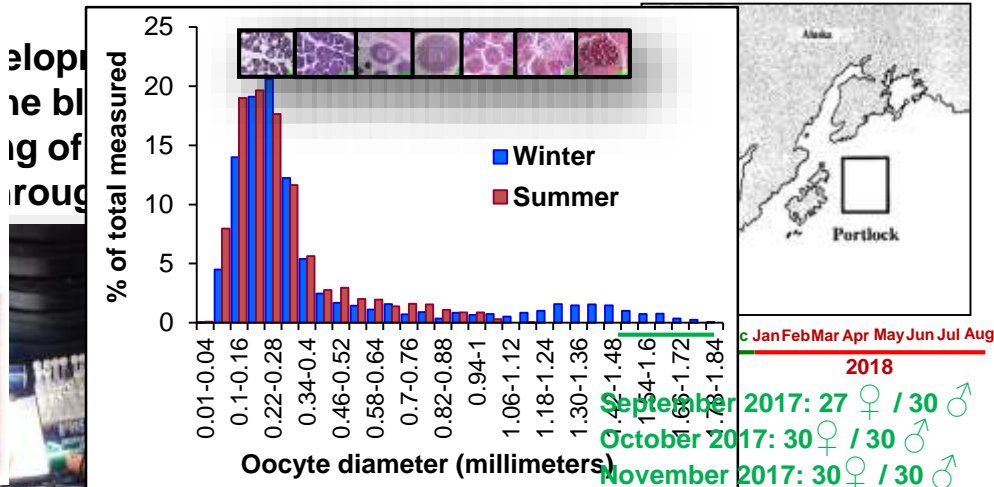
- 71 US vessels
- 16 BC vessels
- Wide participation WA Tribes



# 1. Reproduction

- **Full characterization of the annual reproductive cycle (Project 674.11)**

**Objective:** Understand temporal changes in reproductive development throughout an entire annual reproductive cycle in male and female Pacific halibut



- Comprehensive reproductive development of the adult population in order to improve our estimate of actual spawning biomass



# 1. Reproduction

- Identification of genetic reproductive markers by RNA sequencing

## Ovarian genes

Sample transcript ID	Length (nt)	Database	Database ID	Identity (%)	Gene_symbol	Annotation	Function
TRINITY_DN13531_c0_g1_i1	3754	Danio rerio	ENSDFARP00000004431	89.31	acvr1ba	activin A receptor, type Iba	Oogenesis
TRINITY_DN31883_c0_g1_i1	585	Danio rerio	ENSDFARP000000121689	74.74	adams2	ADAM metalloproteinase with thrombospondin type 1 motif, 2	Ovulation
TRINITY_DN14738_c0_g1_i1	3062	Danio rerio	ENSDFARP000000088795	82.07	ar	androgen receptor	Hormone signaling
TRINITY_DN18096_c0_g1_i1	2654	Danio rerio	ENSDFARP000000076033	76.81	aqp10b	aquaporin 10b	Oocyte hydration
TRINITY_DN18849_c1_g1_i1	1680	Danio rerio	ENSDFARP00000012455	94.97	bmp1a	bone morphogenetic protein 1a	Oogenesis
TRINITY_DN16677_c1_g1_i1	876	Danio rerio	ENSDFARP00000011604	70.78	cyp19a1a	cytochrome P450, family 19, subfamily A, polypeptide 1a	Aromatase (estrogen production)
TRINITY_DN16252_c0_g1_i1	2580	Danio rerio	ENSDFARP00000014026	76.09	dead4	DEAD (Asp-Glu-Ala-Asp) box polypeptide 4	Oogenesis marker
TRINITY_DN23892_c0_g1_i1	232	Danio rerio	ENSDFARP00000073932	71.62	EGFR	epidermal growth factor receptor	Maturation signaling
TRINITY_DN4356_c0_g1_i1	209	Uniprot	QJAK73	66.67	HS017811	estradiol 17-beta-dehydrogenase 11	Steroidogenesis
TRINITY_DN21358_c3_g1_i2	4620	Uniprot	Q9WVM2	80.88	esr2	estrogen receptor beta	Hormone signaling
TRINITY_DN6202_c1_g1_i1	287	Danio rerio	ENSDFARP000000096529	79.79	fshr	follicle stimulating hormone receptor	Hormone signaling
TRINITY_DN9106_c0_g1_i1	2006	Danio rerio	ENSDFARP000000061827	80.39	foxl2	forkhead box L2	Female sex differentiation
TRINITY_DN21868_c0_g1_i1	306	Danio rerio	ENSDFARP00000005566	70.83	gnrh4	gonadotropin releasing hormone receptor 4	Hormone signaling
TRINITY_DN16738_c0_g1_i1	1391	Danio rerio	ENSDFARP000000059732	71.32	inhibb	inhibin, beta B	Oogenesis
TRINITY_DN21305_c0_g1_i1	1466	Uniprot	Q90674	62.65	LHCGR	Lutropin-choriogonadotropin hormone receptor	Hormone signaling
TRINITY_DN12886_c1_g1_i1	771	Danio rerio	ENSDFARP000000109370	82.1	prg	progesterone receptor	Maturation signal
TRINITY_DN15432_c0_g1_i1	2592	Danio rerio	ENSDFARP00000003684	80.26	ptgs2b	prostaglandin-endoperoxide synthase 2b	Prostaglandin synthesis
TRINITY_DN14537_c0_g1_i1	3164	Danio rerio	ENSDFARP00000006091	77.49	mmp2	matrix metalloproteinase 2	Ovulation
TRINITY_DN24972_c0_g1_i1	292	Uniprot	P41243	76.92	Mmp9	Matrix metalloproteinase 9	Ovulation

Oogenesis/oocyte maturation  
Hormone production  
Ovulation



## Testicular genes

Sample transcript ID	Length (nt)	Database	Database ID	Identity (%)	Gene_symbol	Annotation	Function
TRINITY_DN26811_c1_g1_i1	2574	Danio rerio	ENSDFARP000000088795.3	71.15	ar	androgen receptor	Spermatogenesis
TRINITY_DN37544_c0_g1_i1	248	Uniprot	sp C6K1B9 CTSG2_MOUSE	68.75	CatSperp2	Cation channel sperm-associated protein subunit gamma 2	Sperm activation
TRINITY_DN32484_c1_g1_i1	1411	Danio rerio	ENSDFARP000000123870.1	78.57	dead4	DEAD (Asp-Glu-Ala-Asp) box polypeptide 4	PGC marker
TRINITY_DN43322_c0_g1_i1	2573	Uniprot	sp Q80178 DMRT1_CHYL	65.77	dmrt1	Daubmanlike- and mab-3-related transcription factor 1	Male sex differentiation factor
TRINITY_DN23128_c0_g1_i1	1126	Danio rerio	ENSDFARP000000130981.1	85.68	folliculin a	folliculin a	Hormone
TRINITY_DN46346_c0_g1_i1	424	Danio rerio	ENSDFARP000000130239.1	80.56	INHBB	inhibin beta B	Hormone receptor
TRINITY_DN49968_c0_g1_i1	310	Danio rerio	ENSDFARP000000131027.1	79.61	gnrh4	gonadotropin releasing hormone receptor 1	Hormone receptor
TRINITY_DN15629_c0_g2_i1	2899	Danio rerio	ENSDFARP000000088386.3	82.09	nanos3	nanos homolog 3	Spermatogenic marker
TRINITY_DN32333_c1_g1_i1	2783	Danio rerio	ENSDFARP000000109370.2	83.42	prg	progesterone receptor	Hormone receptor
TRINITY_DN6999_c3_g1_i1	345	Danio rerio	ENSDFARP000000104772.2	74.56	ptgs1	prostaglandin-endoperoxide synthase 1	Prostaglandin production
TRINITY_DN20678_c0_g1_i1	124	Danio rerio	ENSDFARP000000136548.1	97.83	RSBN1	round spermatid basic protein 1	Spermatid marker
TRINITY_DN33366_c1_g1_i1	3258	Danio rerio	ENSDFARP000000104616.2	89.27	stbp	spermatid perinuclear RNA binding protein	Spermatid marker
TRINITY_DN34579_c8_g1_i1	635	Danio rerio	ENSDFARP000000106978.2	92.67	sax3a	SRY (sex determining region Y)-box 3a	Male sex differentiation factor
TRINITY_DN6843_c0_g1_i1	235	Danio rerio	ENSDFARP00000021907.6	79.49	star	steroidogenic acute regulatory protein	Testicular steroidogenesis

Spermatogenesis  
Testicular differentiation  
Sperm production



Halibut gonadal transcriptome dataset



# Primary research areas at IPHC



1. Reproduction

2. **Growth**

- CHANGES IN SIZE AT AGE/BIOMASS
- TOOLS TO ASSESS FISH CONDITION

3. DMRs and post-release survival assessment

4. Migration

5. Genetics and genomics



## 2. Growth

Little is known regarding what factors influence growth in this species

- CHANGES IN SIZE AT AGE/BIOMASS
- TOOLS TO ASSESS FISH CONDITION

### Projects:

1. *Identification and validation of physiological markers for growth (Project 673.14)*
2. *Evaluation of growth patterns and effects of environmental influences (NPRB 1704)*

### Objectives:

- Knowledge on growth patterns and environmental influences.
- Improved understanding in the possible role of growth alterations in the observed decrease in size at age.



# 2. Growth

## 1. Mass identification

**Objective:** Identify

- Identification of growth-related genes
- Develop molecular assays

Annotation	Gene symbol	Length (nt)	Identity (%)	Function
Androgen receptor	<i>ar</i>	4426	81.48	Protein synthesis
Calcium/calmodulin-dependent protein kinase II alpha	<i>camk2a</i>	2342	87.27	Force transmission
Creatine kinase, muscle a	<i>ckma</i>	2256	89.76	Energy metabolism
Carnitine palmitoyltransferase 1B	<i>cpt1b</i>	762	81.81	Lipid metabolism
Dystrophin	<i>dmd</i>	1282	82.23	Force transmission
Eukaryotic translation initiation factor 4eb	<i>eif4eb</i>	1168	85.19	Protein synthesis
F-box protein 32	<i>fbxo32</i>	695	86.25	Protein atrophy
Glycogen synthase 1	<i>gys1</i>	3723	89.47	Energy metabolism
Histone deacetylase 1	<i>hdac1</i>	2490	96.15	Muscle repressor
Insulin-like growth factor 2 receptor	<i>igf2r</i>	511	80.52	Growth regulator
Insulin-like growth factor binding protein 5b	<i>igfbp5</i>	1372	81.5	Growth regulator
Lipoprotein lipase	<i>lln</i>	1789	60.48	Lipid metabolism
Myocyte enhancer factor 2cb	<i>mef2cb</i>	564	79.8	Muscle growth
Myostatin b	<i>mstnb</i>	189	95.71	Growth regulator
Mechanistic target of rapamycin	<i>mtor</i>	1153	97.92	Protein synthesis
Myogenic factor 6	<i>myf6</i>	819	81.19	Muscle growth
Myosin, heavy polypeptide 1.3, skeletal muscle	<i>myh13</i>	246	86.42	Muscle growth
Myoblast determination protein 1 homolog	<i>myod</i>	2407	72.67	Muscle development
Myozenin 1a	<i>myoz1a</i>	766	74.6	Force transmission
Nuclear factor of activated T-cells, cytoplasmic 3	<i>nfatc3</i>	1587	62.96	Muscle activity
Paired box 3a	<i>pax3</i>	269	75	Muscle development
Paired box 7b	<i>pax7b</i>	297	85.71	Muscle development
Peroxisome proliferator-activated receptor gamma, coactivator 1 alpha	<i>ppargc1a</i>	519	88.7	Energy metabolism
Protein phosphatase 3, catalytic subunit, alpha isozyme	<i>ppp3ca</i>	3407	83.69	Muscle activity
Protein kinase, AMP-activated, alpha 1 catalytic subunit	<i>prkaa1</i>	1925	70.96	Energy metabolism
Phosphorylase, glycogen, muscle	<i>pygma</i>	5514	90.91	Energy metabolism
Serum response factor	<i>srf</i>	4393	63.81	Muscle development
Transforming growth factor, beta 1a	<i>tgfb1a</i>	561	77.04	Growth regulator
Tripartite motif containing 63b	<i>trim63b</i>	2117	81.16	Protein atrophy

**Deliverables:**

- Establishment of a growth-related gene sequence dataset
- Molecular assays to monitor growth patterns based on growth-markers



# 2. Growth

## 2. Evaluation of growth patterns and effects of environmental influences

**Objective:** Identify molecular, biochemical and isotopic profiles characteristic of specific growth patterns and evaluate potential effects of environmental influences.

- Establishment of different growth trajectories in juvenile fish in captivity to identify molecular and biochemical signatures of growth patterns.
  - *Manipulating growth rates (ration, density, thermal- or fasting-induced compensation, etc.):*



- Evaluation of different growth patterns in the *wild*.

Samples collected in NMFS trawl survey  
In 2016 and 2017 from 3 size categories:

- <40 cm length
- 40-60 cm length
- 60-80 cm length



*Characterization of molecular and biochemical growth markers in muscle samples from age-matched individuals*

- Isotopic tissue turnover to trace dietary and/or habitat shifts

$^{13}\text{C}$ ,  $^{15}\text{N}$



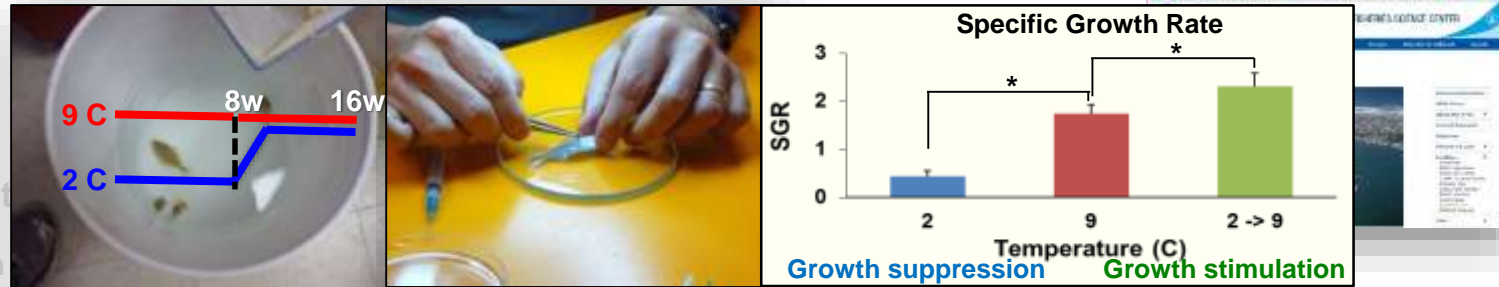


## 2. Growth

- **Evaluation of growth patterns and effects of environmental influences**

**Objective:** Identify molecular, biochemical and isotopic profiles characteristic of specific growth patterns and evaluate potential effects of environmental influences.

- Establishment of different growth trajectories in juvenile fish in captivity to identify molecular and biochemical signatures of growth patterns.



- 75 fish <40 cm length
  - 75 fish 40-60 cm length
  - 75 fish 60-80 cm length
- Continued in 2017

growth markers in muscle samples from age-matched individuals

- Isotopic tissue turnover to trace dietary and/or habitat shifts

$^{13}\text{C}$ ,  $^{15}\text{N}$

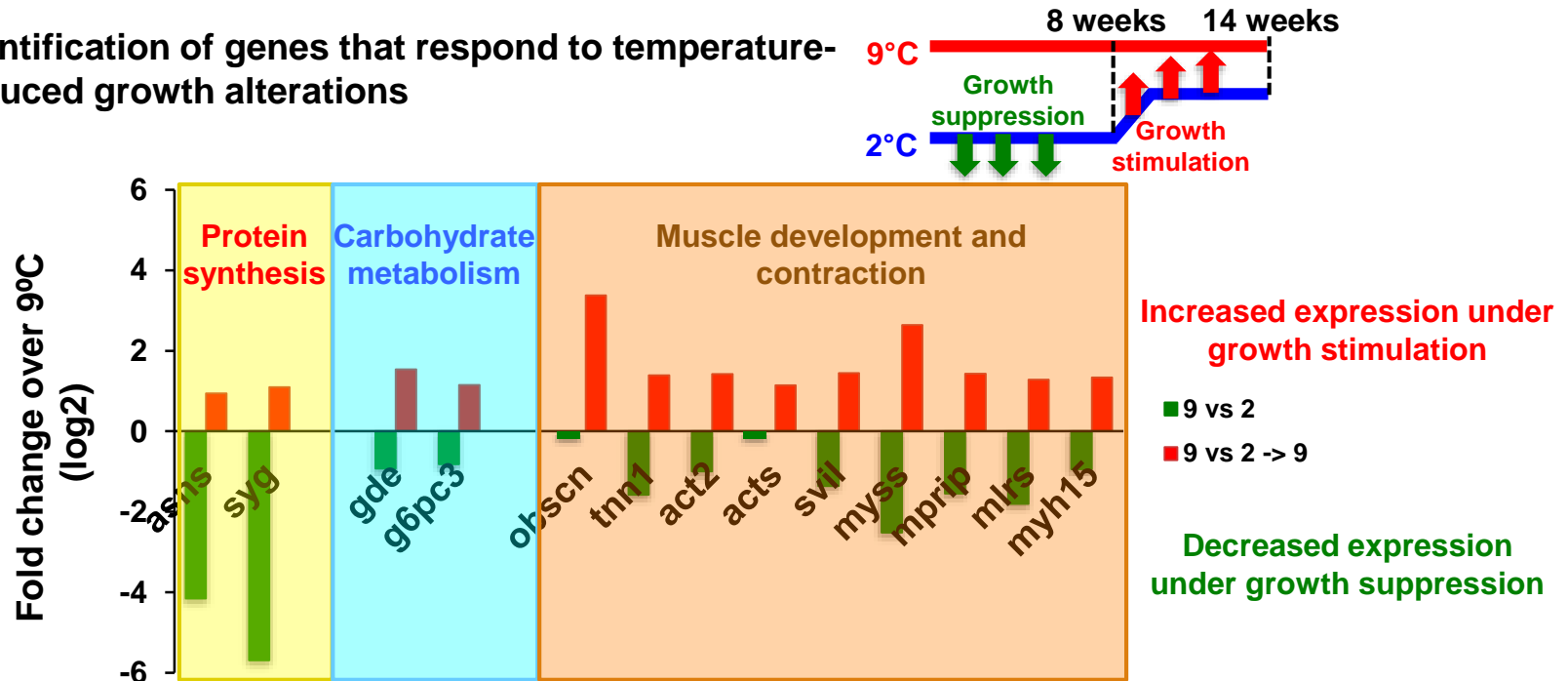




## 2. Growth

- Evaluation of growth patterns and effects of environmental influences*

Identification of genes that respond to temperature-induced growth alterations



Potential molecular markers for temperature-regulated growth



## 2. Growth

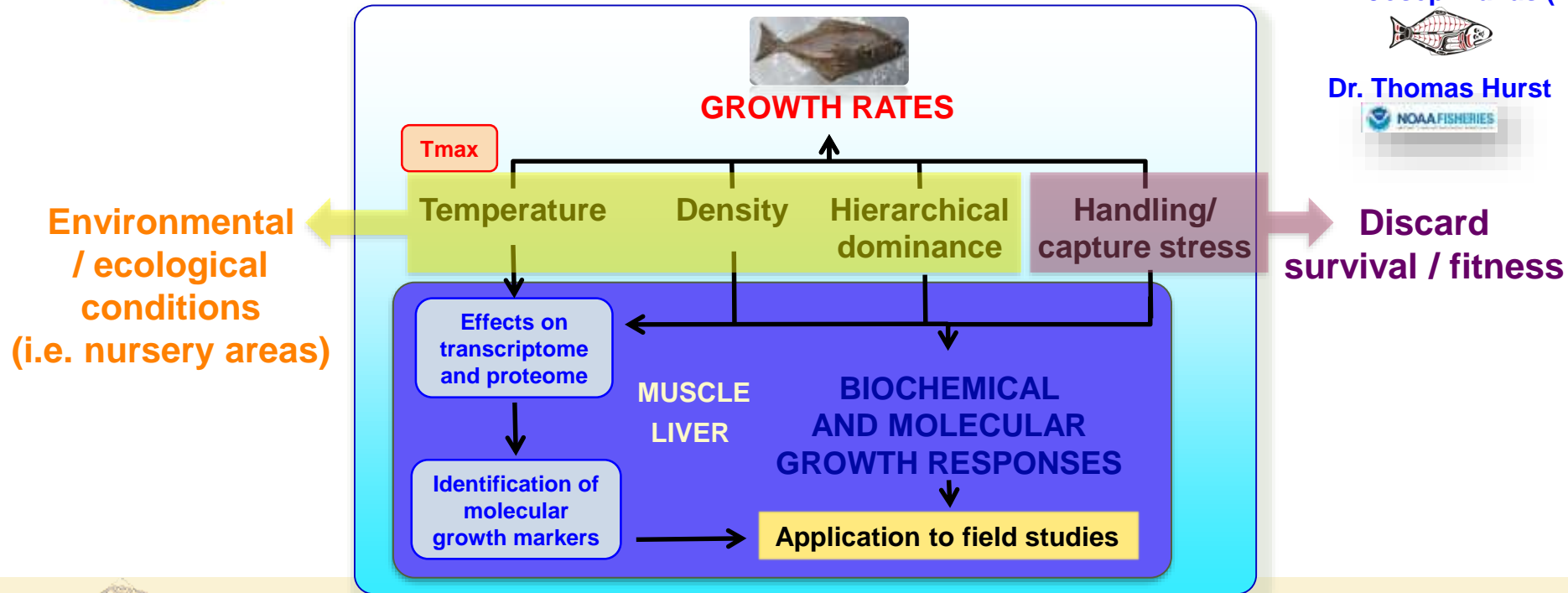


- NPRB Grant 1704 (2017-2019):** “Somatic growth processes in the Pacific halibut (*Hippoglossus stenolepis*) and their response to temperature, density and stress manipulation effects”. IPHC / AFSC – Newport, OR

Dr. Josep Planas (PI)



Dr. Thomas Hurst



## 2. Growth

- **Evaluation of growth patterns and effects of environmental influences**

**Objective:** Identify molecular, biochemical and isotopic profiles characteristic of specific growth patterns and evaluate potential effects of environmental influences.

- Evaluation of different growth patterns in the *wild*.

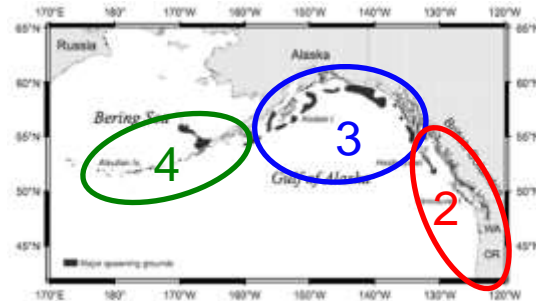
Samples collected in NMFS trawl survey  
In 2016 and 2017 from 3 size categories:

- <40 cm length
- 40-60 cm length
- 60-80 cm length



*Characterization of molecular and biochemical growth markers in muscle samples from age-matched individuals*

- Phase 2: Regional monitorization of growth patterns



## 2. Growth

- Investigate the effects of other **environmental factors** on growth performance.
  - Effects of **salinity**, **dissolved oxygen** and **water pH** on growth.



*Relate to catch efforts in FISS in time and space model*

- *Identify the optimal environmental conditions for growth.*

### Deliverables:

- Identification and validation of growth markers for field studies
- Characterization of molecular and biochemical growth signatures
- Environmental effects on somatic growth
- Improved biological inputs on biomass estimates



# Primary research areas at IPHC



1. Reproduction

2. Growth

3. DMRs and post-release survival assessment

• BYCATCH SURVIVAL  
ESTIMATES

4. Migration

5. Genetics and genomics



# 3. DMRs and survival assessment

Little is known regarding the factors that influence bycatch survival

- **BYCATCH SURVIVAL ESTIMATES**

## Project components:

- 1. Evaluate the effects of fish handling practices on injury levels and the physiological condition of captured Pacific halibut (Project 672.13, S-K Grant)*
- 2. Investigate the relationship between physiological condition post-capture and survival as assessed by the use of accelerometer tags (S-K Grant)*
- 3. Explore the applicability of electronic monitoring in DMR estimations (S-K Grant)*

## Objectives:

- To introduce quantitative measurable factors that are linked to fish handling practices and to fish physiological condition and ultimately to survival in order to improve current DMR estimations



### 3. DMRs and survival assessment

- *Evaluate the effects of fish handling practices on injury levels and the physiological condition of captured Pacific halibut*

**Objective:** Understand relationship between handling practices and physiological condition of captured Pacific halibut in the longline fishery

- Assess **injuries** associated with release techniques (gangion cut, careful shake, hook straightening, hook stripping).



### 3. DMRs and survival assessment

- *Investigate the relationship between physiological condition post-capture and survival as assessed by tagging*

**Objective:** Measure post-release survival in Pacific halibut and relate it to physiological condition and capture-related events

- Tag fish that have been exposed to different handling practices in the longline fishery with accelerometer tags in addition to conventional tags (wire).
- Assess survival of fish according to size and physiological condition.

Deliverables

- Information on physiological condition
- Information on survival





### 3. DMRs and survival assessment

- *Explore the applicability of electronic monitoring in DMR estimations*

**Objective:** Test the ability of electronic monitoring to capture fish handling events and fish condition and relate it to survival

- Deploy electronic monitoring (EM) system on a longline vessel.
- Video record fish handling events during capture.
- Determine injury profile by release method.



Deliver

- Determine injury profile by release method and associated injury levels and projected survival.

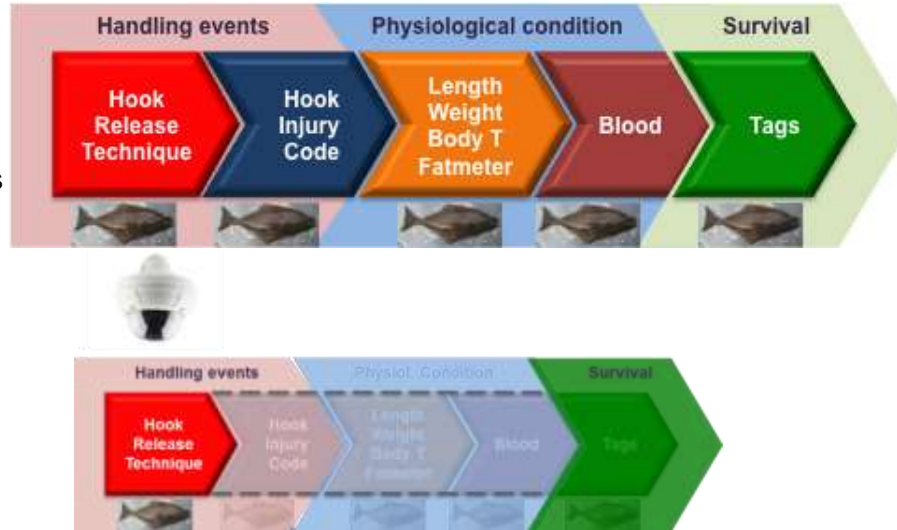


# 3. DMRs and survival assessment

- **Saltonstall – Kennedy Grant NA17NMF4270240 (2017-2019):** “*Improving discard mortality rate estimates in the Pacific halibut by integrating handling practices, physiological condition and post-release survival*”. IPHC / APU – Anchorage, AK

## November 2017

- 2 6-day trips (GOA, F/V Kema Sue)
- 38 sets (8 standard skates/set)
- 3 randomized treatments/skate
- 1,048 fish sampled and wire tagged
- 79 fish tagged with accelerometer tags (mini satellite tags; 96 days recording)
- EM on each haul



Dr. Josep Planas (PI)  
Claude Dykstra  
Dr. Tim Loher  
Dr. Ian Stewart  
Dr. Allan Hicks



Dr. Brad Harris  
Dr. Nathan Wolf



# Primary research areas at IPHC



1. Reproduction
2. Growth
3. DMRs and post-release survival assessment
4. **Migration**
  - ADULT FEEDING AND REPRODUCTIVE MIGRATION
  - LARVAL DISPERSAL
5. Genetics and genomics



# 4. Migration

- ADULT FEEDING AND REPRODUCTIVE MIGRATION
- LARVAL DISPERSAL

## Projects:

1. *Juvenile and adult feeding migrations (Project 670.11)*
2. *Tail pattern recognition (Project 675.11)*
3. *Adult dispersal on Bowers Ridge (Reg. Area 4B) (Project 650.21)*
4. *Larval migration and connectivity*

## Objectives:

- To improve our understanding on larval, juvenile and reproductive migration.



# 4. Migration

- ***Juvenile and adult migration studies (Project 670.11)***

- Juvenile wire tagging:
  - NMFS trawl tagging project: **1469 fish**
    - 713 fish in GOA and 756 fish in BS
- Adult wire tagging:
  - IPHC survey tagging project
    - 2016 pilot study in area 4D (U32)
    - 2017 coast-wide study (U32): **1927 fish**

**Fin clips** are collected: Genetic analyses of tagged fish to shed light on migration patterns and geographic origin.



- ***Tail pattern recognition (Project 675.11)***



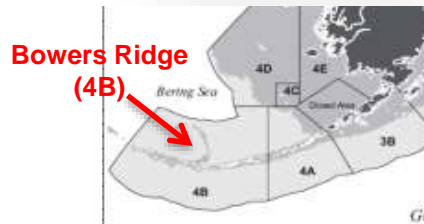
- Blind side of tail is preferable for imaging.
- Spots and patterns appear to be unique.
- Tail markings can be used to identify individuals with image recognition software.
- Promising for implementation in FISS.



# 4. Migration

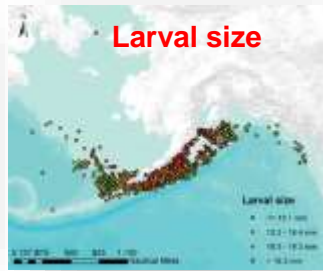
- ***Reproductive and annual migration (Project 650.21)***

- In 2017: 14 adult fish tagged with sPAT tags (12 females, 2 males). No tag information returned.



- ***Larval migration and connectivity***

**Objective:** Understand the mechanisms of larval connectivity between GOA and BS.



Collaboration with Janet Duffy-Anderson,  
Esther Goldstein,  
William Stockhausen (NOAA-AFSC-Seattle)



# Primary research areas at IPHC



1. Reproduction
2. Growth
3. DMRs and post-release survival assessment
4. Migration
5. **Genetics and genomics**
  - GENETIC STRUCTURE OF THE POPULATION
  - GENOMIC TOOLS (e.g. GENOME)



# 5. Genetics and genomics

- GENETIC STRUCTURE OF THE POPULATION
- GENOMIC TOOLS (e.g. GENOME)

## Projects:

- 1. Sequencing of the Pacific halibut genome (Project 673.13)*
- 2. Population genetic studies*

## Objectives:

- Improve knowledge on the genetic composition of the population
- Establish genomic resources for the species
- Evaluate effects of fishery-dependent and fishery-independent influences on growth, reproduction, nutrition, etc.





# 5. Genetics and genomics

- *Pacific halibut genome sequencing (Project 673.13)*



**Objective:** Generate a first draft sequence of the Pacific halibut genome

- Genomic DNA sequenced from one Pacific halibut female (WZ).
- Conducted first genome assembly:
  - Full genome sequenced. Genome size: 700 Mb
  - Non-continuous genome sequence.
- Additional sequencing is required to complete assembly.



Dr. Yann Guiguen



Dr. Lorenz Hauser



# 5. Genetics and genomics

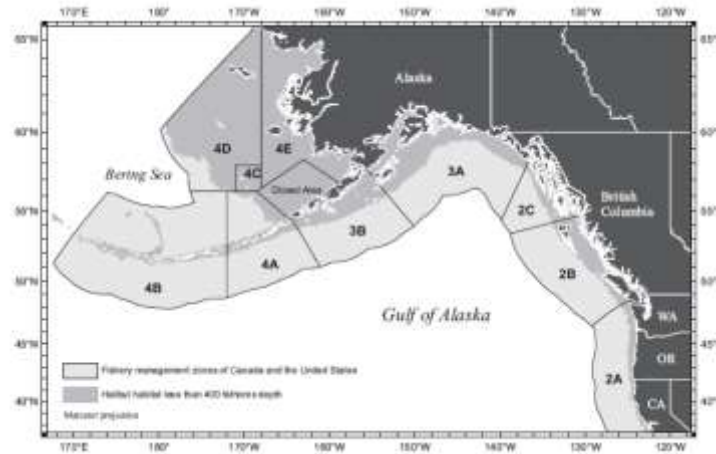
- *Population genetic studies (PLANNED)*

**Objective:** Genetic characterization of Pacific halibut throughout its distribution range

- Characterization of population structure by RAD sequencing and SNP analysis.
- Identification of possible genetic signatures of geographical origin



Dr. Lorenz Hauser



# Outline of the presentation



- Update on the research activities of the Biological and Ecosystem Science Program
- **Outcome of external funding applications**
- Proposed research projects for 2018
- Establishment of the IPHC research laboratory



# Research proposals submitted for external funding in 2017

Project #	Grant agency	Project name	Partners	IPHC Budget (US\$)	PI	Management implications	Submission status
1	<b>Saltonstall-Kennedy NOAA</b>	Improving discard mortality rate estimates in the Pacific halibut by integrating handling practices, physiological condition and post-release survival	Alaska Pacific University	223,220	Planas (lead PI) Dykstra Loher Stewart Hicks	Bycatch estimates	Awarded
2	<b>NPRB</b>	Somatic growth processes in the Pacific halibut ( <i>Hippoglossus stenolepis</i> ) and their response to temperature, density and stress manipulation effects	AFSC-NOAA-Newport	122,264	Planas (lead PI)	Changes in biomass/size-at-age	Awarded
3	<b>NPRB</b>	Larval transport, supply, and connectivity of Pacific halibut between the Gulf of Alaska and the Bering Sea	AFSC-NOAA-Seattle UAF	8,000	Sadorus Planas Stewart	Biomass distribution	Rejected
4	<b>Essential Fish Habitat NOAA</b>	Validating biochemical markers of growth for habitat assessment in flatfishes	AFSC-NOAA-Newport	35,000	Hurst (lead PI) Planas	Changes in biomass/recruitment	Rejected
5	<b>NFWF</b>	Evaluating virtual vitality assessments of discarded Pacific halibut	AFSC-NOAA, APU, NFR	-	Harris (APU), Dykstra	Bycatch estimates	Rejected



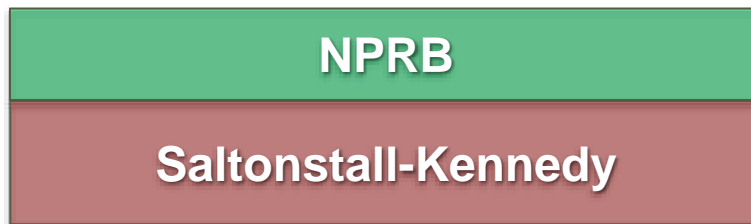
# Outcome of external funding applications

Project #	Grant agency	Project name	Partners	IPHC Budget (\$US)	PI/IPHC Staff	Management implications	Submission status
1	Saltonstall-Kennedy NOAA	Improving discard mortality rate estimates in the Pacific halibut by integrating handling practices, physiological condition and post-release survival	Alaska Pacific University	\$223,220	Planas (lead PI) Dykstra Loher Stewart Hicks	Bycatch estimates	<b>Awarded</b>  Started in September 2017
2	NPRB	Somatic growth processes in the Pacific halibut ( <i>Hippoglossus stenolepis</i> ) and their response to temperature, density and stress manipulation effects	AFSC-NOAA-Newport	\$131,891	Planas (lead PI) Rudy	Changes in biomass/size-at-age	<b>Awarded</b>  Started in September 2017
Total awarded (\$US)				\$355,111			



# Temporal chart of activities

	2016	2017	2018	2019	2020	2021
Reproduction		Annual reproductive cycle				
			Sex determination mechanisms			
	Sex identification					



# Outline of the presentation



- Update on the research activities of the Biological and Ecosystem Science Program
- Outcome of external funding applications
- **Proposed research projects for 2018**
- Establishment of the IPHC research laboratory



# Research projects proposed for 2018

Project #	Project Name	Priority	Budget (\$US)	External funding for FY2018 (\$US)	Management implications
<b>New Projects</b>					
2018-01	Influence of thermal history on growth	High	\$136,004	-	Changes in biomass / size-at-age
2018-02	Adult captive holding studies	High	\$53,395	-	Changes in biomass / size-at-age / larval distribution
2018-03	Whale detection methods	High	\$37,511	-	Mortality estimation
2018-04	Larval connectivity modeling	High	\$20,000	-	Larval distribution
<b>Continuing Projects</b>					
621.16	Development of genetic sexing techniques	High	\$33,928	-	Sex composition of the catch
642.00	Assessment of Mercury and other contaminants	Medium	\$8,400	-	Environmental effects
650.18	Archival tags: tag attachment protocols	High	\$800	-	Adult distribution
650.21	Investigation of Pacific halibut dispersal in Regulatory Area 4B	High	\$6,800	-	Spawning areas
661.11	Ichthyophonous Incidence Monitoring	Medium	\$8,755	-	Environmental effects
669.11	At-sea Collection of Pacific Halibut Weight to Reevaluate Conversion Factors	High	\$7,645	-	Length-weight relationship
670.11	Wire tagging of Pacific halibut on NMFS trawl and setline surveys	High	\$12,840	-	Juvenile and adult distribution
672.12	Condition factors for tagged U32 Fish	High	\$9,116	-	DMR estimates
672.13	Discard mortality rates and injury classification profile by release method	High	\$1,037	\$255,402	DMR estimates
673.13	Sequencing the Pacific halibut genome	High	\$32,500	-	Environmental/Fishery effects
673.14	Identification and validation of markers for growth	High	\$25,681	\$57,773	Changes in biomass / size-at-age
674.11	Full characterization of the annual reproductive cycle	High	\$121,488	-	Maturity assessment
675.11	Tail pattern recognition	High	\$3,900	-	Juvenile and adult distribution
	Total - New Projects		\$251,910		
	Total - Continuing Projects		\$273,090		
	Overall Total (all projects for FY2018)		\$525,000		
	External Funding (for FY2018) (\$US)			\$313,175	



# New research projects proposed for 2018

- **Influence of thermal history on growth**

- Relate temperature history to individual growth as assessed by archival tagging
- Tag U32 fish with electronic archival tags recording temperature and depth.
- Quantify growth patterns in captured fish and relate them to the thermal history.
- Compare archival data analyses with otolith microchemistry ( $O^{18}$ ).

- **Whale de**

- Test ac
- whale de
- Relate w
- captures



ologies for  
cific halibut



- **Adult captive holding studies**

- Test permanence of individual tail markings (Tail Pattern Recognition)
- Conduct diet manipulation experiments: fat meter validation, stable isotope studies on growth ( $N^{15}/C^{13}$ )
- Conduct temperature manipulation experiments for growth and  $O^{18}$  calibration studies
- Perform larval swimming performance tests
- Test transgenerational marking approaches through broodstock labeling



- **Larval co**

- Model larval abundance and size distribution in the GOA and BS over time and oceanographic and environmental conditions.



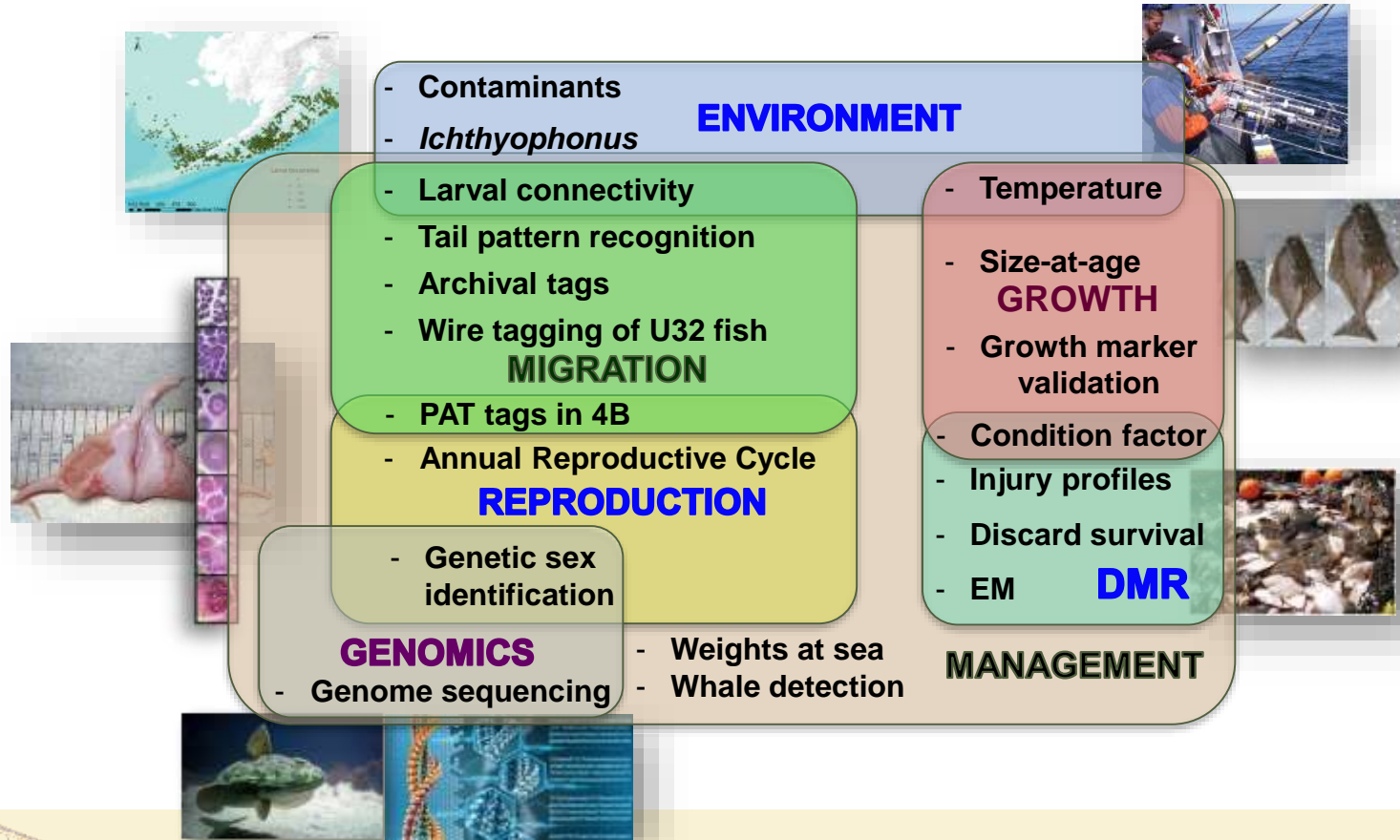
Warm stanza



Cold stanza



# Research projects for 2018



# Outline of the presentation



- Update on the research activities of the Biological and Ecosystem Science Program
- Outcome of external funding applications
- Proposed research projects for 2018
- **Establishment of the IPHC research laboratory**



# IPHC Research Laboratory



- **Laboratory space has been remodeled**
- **Laboratory equipment is being purchased**
- **A laboratory technical position is requested**
- **Biological samples will be processed and analyzed in house:**
  - **Genetic sex assays: genotyping**
  - **Growth and reproductive assessment**
  - **Blood hormone levels**
  - **Stress and disturbance indicators**



